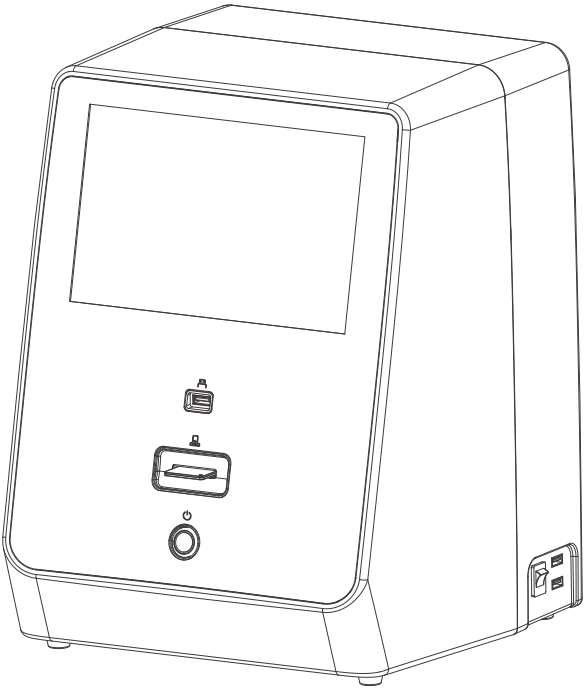
Servicebio



User Manual

Fluorescence Cell Counter

SCC-F1000

Please read the User Manual carefully and keep it properly before using the product for future reference.

Wuhan Servicebio Technology Co.,Ltd.

CONTENTS

01 Product Description & Precautions

Product Description

Thank you for purchasing our product. To fully enjoy our company's comprehensive service, please read the instructions and related illustrations carefully before using this product.

The fluorescence cell counter is a precise and fast cell analysis system that combines image recognition technology and optical imaging technology to obtain data such as cell number, concentration, and viability with one click, and display the morphology of cells.

Danger/Warning

1. Never place the power cord or plug in water or any other liquid;

2. Malfunction or damage caused by not following the instructions will not be covered by the warranty.

Precautions

1. Always place the fluorescence cell counter on a flat, stable, heat-resistant workbench;

2.Do not use in environments with visible water, excessive moisture, or near open flames and extreme heat;

3.If the fluorescence cell counter cannot operate normally, unplug the power plug immediately;

4.Do not pull the power cord, and be careful when unplugging the power plug;

5.Do not move the fluorescence cell counter while it is working;

6.The fluorescence cell counter should be protected against moisture and freezing;

7.If not in use for a long time, the power switch should be turned off and the power plug should be unplugged;

8.Do not use the fluorescence cell counter if the power cord or plug is damaged. Please contact our company's service center for replacement or repair;

9.Damage caused by not following the instructions is not covered by the warranty.

02 Technical Parameters

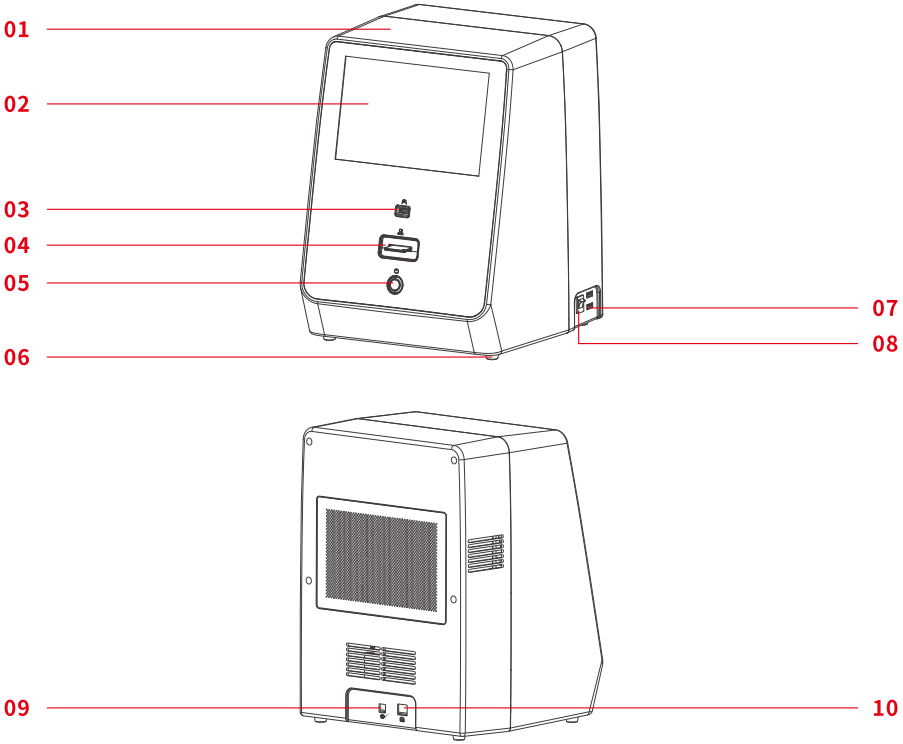
Product Parameters

Product Name	Fluorescence Cell Counter
Cat. No.	SCC-F1000
Dimensions	287×283×388mm
Net Weight	12Kg
Electrical Parameters	Input 100~240VAC 50/60Hz,4.0A Output 12V,15A,180W
Operating Environment	10°C~40°C, Relative Humidity <80%
Consumable Type	Disposable Counting Chamber
Consumable Throughput	6-Channel
Counting Modes	Trypan Blue Staining Analysis / Unstained Analysis / AO/PI Staining Analysis / GFP Transfection Analysis
Focusing Method	Auto-focus
Camera Pixel	11 Megapixel CMOS
Optical Magnification	10×
Light Source	Green Fluorescence Ex480±20nm, Em530±25nm; Red Fluorescence Ex530±20nm, Em620±30nm; White LED
Field of View Image	Single Channel with 4 Views of Photography
Display Module Resolution	10.1",1280×800
Image Resolution	4128×2808
Image Format	JPG

Result Output Format	JPG/PDF/CSV
Storage Space	256G
Storage Records	1000 pcs
Sample Types	Suspension Culture Cells / Adherent Culture Cells / Mammalian Cells / Human-derived Cells / PBMC, etc.
Concentration Range	5×10^4~1.5×10^7 cells/mL (Optimal5×10^5~1×10^7 cells/mL)
Diameter Range	2~180μm (Optimal: 2~180μm)
Counting Time	Approx. 320s for Dual Fluorescence All Channels
Sample Volume	10μL

03 Product Overview

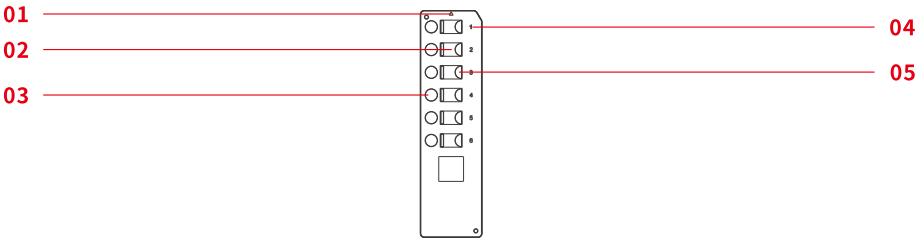
Product Structure Description



01 Main Body	02 Screen	03 USB 3.0 Port
04 Cell Counting Chamber Connector	05 Switch	06 Rubber Foot Pad
07 USB 2.0 Port	08 Power Switch	09 Network Port
10 DC Power Port		

04 Product Operation

4.1 Sample Preparation



Cell Counting Chamber

01 Insertion Direction	02 Sample Chamber	03 Mixing Pool
04 Well Number	05 Loading Area	

1.Take out our company's cell counting chamber. As shown in the image above, at the 03 position (mixing chamber) on the left side of the counting chamber, uniformly mix the cell suspension with the dedicated staining solution (Trypan Blue staining solution or AO/PI staining solution) according to a specified ratio to prepare the sample for testing;

2.Using a pipette, gently transfer 10 μL of the test sample to the semi-circular loading area on the right side of the counting chamber. The sample will spread evenly across each chamber via capillary action;

3.After completing the sample addition, allow the counting chamber containing the samples to rest for 30 seconds. Then, insert the counting chamber horizontally into the loading port at the front of the machine, pushing it all the way to the end.

4.1.1 Mixing and Staining Methods Introduction

1.Trypan Blue Staining Analysis: Prepare the test sample by mixing the cell suspension with our company's Trypan Blue stain(ready-to-use) (0.2%) in a 1:1 ratio. Using a pipette, gently dispense 10 μL of the test sample into the counting chamber. Allow the sample to settle before proceeding with machine detection;

2.AO/PI Staining Analysis: Prepare the test sample by mixing the cell suspension with our company's AO/PI stain(ready-to-use) in a 1:1 ratio. Using a pipette, gently dispense 10μL of the test sample into the counting chamber. Allow it to settle before proceeding with machine detection (the device's default exposure parameters are optimised for compatibility with our company's AO/PI staining solution);

3.Unstained Analysis: Pipette 10μL of the cell suspension to be examined into the chamber of the hemocytometer. Allow the sample to settle before proceeding with the analysis;

4.GFP Transfection Analysis: Using a pipette, gently transfer 10 μL of the cell suspension to be examined into the counting chamber. Allow the sample to settle before proceeding with the assay.

Note

1.Avoid pressing or shaking the counting chamber vigorously after adding the sample;

2.When inserting the counting chamber, pay attention to the correct orientation;

3.The cell counting chamber is successfully inserted when it cannot be pushed any further after being pushed into the port;

4.Do not insert the cell counting chamber during the reset and self-check process. During the self-check, a pop-up window with the message "Self-checking" will appear on the screen, and the power button on the front of the instrument will flash red. After the self-check is completed, the green light will remain on, and the pop-up prompt on the screen will disappear;

5.For the Trypan Blue staining analysis function of this equipment, it is recommended to use Servicebio's Trypan Blue Stain(Ready-to-use) (0.2%) for staining cell samples;

6.For the AO/PI staining analysis function of this equipment, it is recommended to use our company's ready-to-use AO/PI dual-stain reagent for staining cell samples;

7.The left side of the cell counting chamber is equipped with six sample mixing pools, each holding 20μL. Residual liquid should be wiped away using tissue paper.

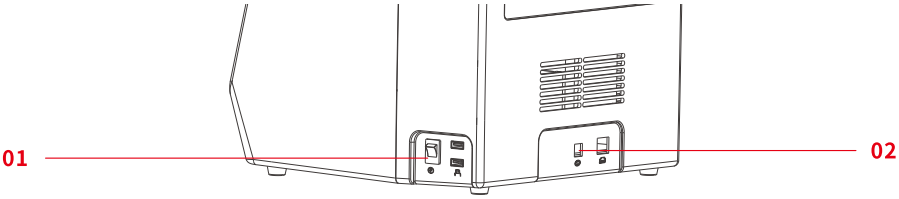
4.2 Power On and Startup

1. Plug the power cable into the DC power port of the instrument;

2. Insert the power cable three-prong plug into an AC power outlet;

3. Press the "I" rocker switch located at the lower right side of the machine to turn on the power. The switch light will remain on;

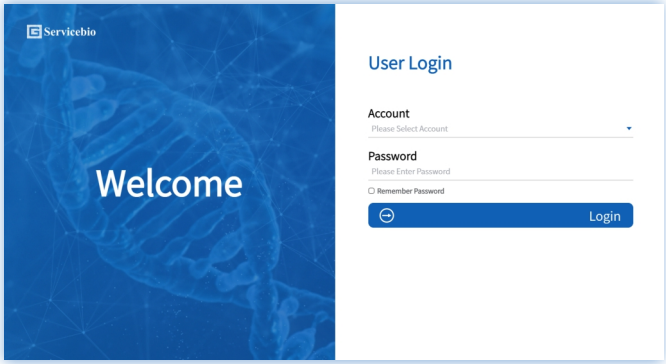
4. Press the power button located below the screen to turn on the device. During the startup process, the button flashing red light indicates that the system is performing a self-check. After the self-check is completed, the green light will remain on.



01 Power Switch

02 DC Power Port

4.3 User Login



Login Interface

Do not perform any operations while the screen shows "Self-testing". After the self-test is complete, a user login window will pop up on the screen. For initial login, fill in the account information, password, check "Remember Password", and log in (initial account: user, initial password: 123456). For subsequent logins, click ▼ to pop up a drop down menu, select the account, and log in(When the device is already logged in and Remember Password is checked, subsequent startup skips the login screen and enters the cell counting main interface automatically).

4.4 Sample Measurement

After login is complete, enter the main interface of the fluorescence cell counter. The system automatically enters the sample measurement interface.

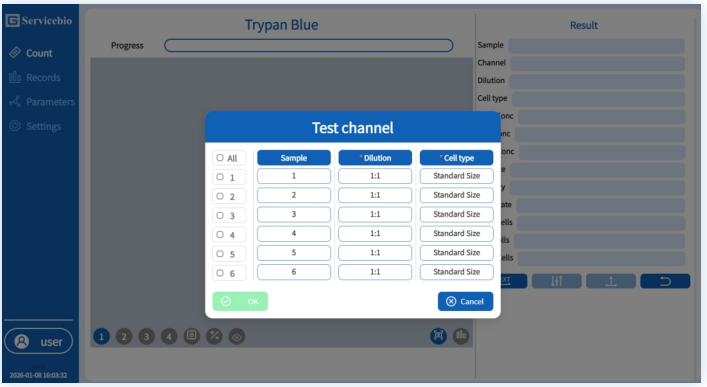
Sample Measurement Interface

The sample measurement interface includes 4 analysis modes: Trypan Blue Staining Analysis / Unstained Analysis / AO/PI Staining Analysis / GFP Transfection Analysis. Users can select the appropriate analysis mode for cell counting based on experimental properties and testing requirements.

The left navigation bar of the fluorescence cell counter interface displays four functional modules: Count Records Parameters Settings. Users may click to view or perform other operations.

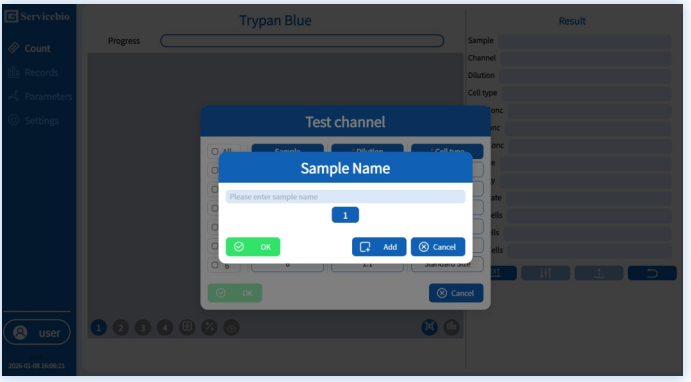
4.4.1 Trypan Blue Staining Analysis

Select the Sample Measurement function, click the Trypan Blue Staining Analysis module on the main interface. The main window will pop up the test well selection interface for Trypan Blue staining analysis. Click Select All to simultaneously select all 6 channels for testing, or select a single channel individually. Choose or create a sample name as needed (default sample name is the channel number). Select the dilution ratio (default is 1:1). Select the cell type (default cell type is standard particles).



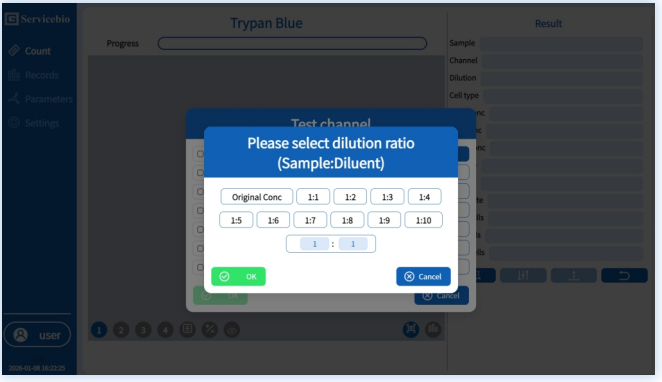
Trypan Blue Staining Analysis/Selcet Test Well Positions Interface

For example, when analyzing channel 1 of the counting chamber, click "1" below the "Sample Name" column for channel 1 to create or select a sample name, defaulting to the channel number.



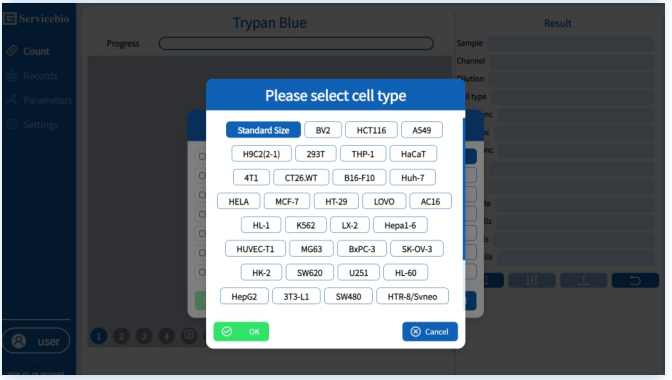
Trypan Blue Staining Analysis/Select Sample Name Interface

Click "1:1" below the Dilution Ratio column to select different dilution ratios. Default is 1:1.



Trypan Blue Staining Analysis/ Select Dilution Ratio Interface

Click Standard Particles below Cell Type to select the cell type. Default is Standard Particles.



Trypan Blue Staining Analysis/ Select Cell Type Interface

After entering the Trypan Blue staining analysis interface, the user should insert the cell counting chamber containing the sample into the port and push it into place, then select the test well positions for measurement.

Click the well position number selection box to choose the wells to be measured, then sequentially select the sample name, dilution ratio, and cell type. You can keep the current well editing status as default, or manually click the knob corresponding to the well to change the editing in bulk or individually. After editing is complete, click OK.

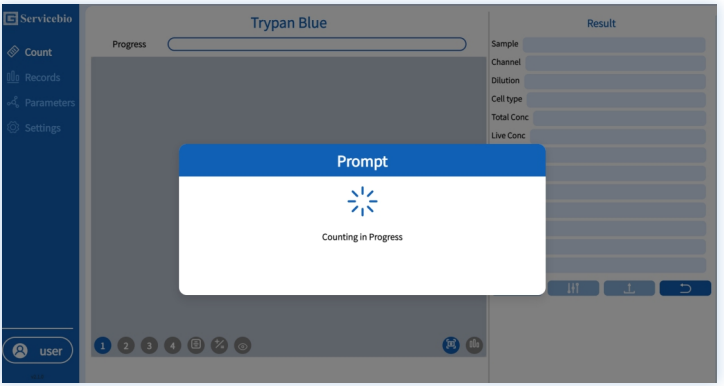
Click NEXT to proceed to the next counting.

Note

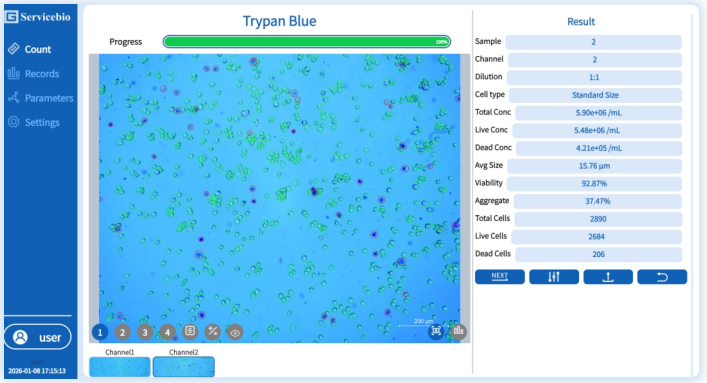
1.The instrument has a total of six test wells. A maximum of six samples can be detected at once. Well position numbers are represented by Arabic numerals 1, 2, 3, 4, 5, 6;

2.The sample name, dilution ratio, and cell type on the select test well position interface are initially set to default. Users can click the display box to edit as needed.

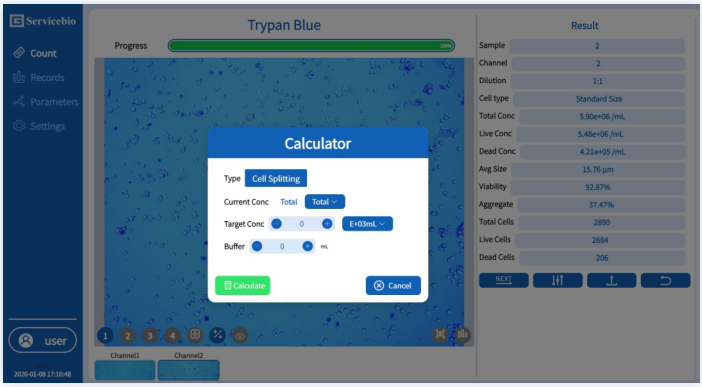
After editing the sample name, dilution ratio, and cell type, click OK to start the Trypan Blue staining counting analysis. The interface is shown below. A "Counting in progress" prompt box will pop up. Do not perform any operations at this time; wait quietly for the counting to complete.



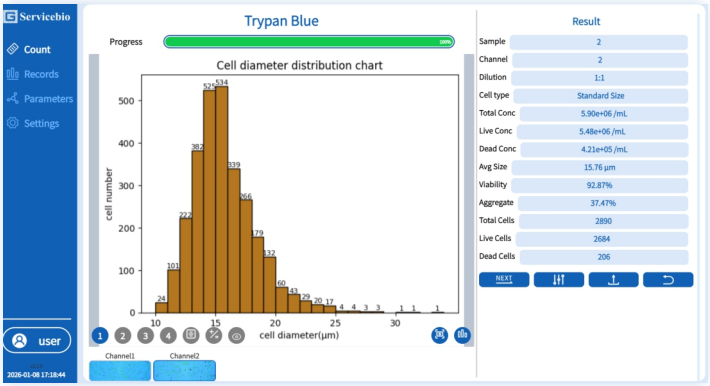
Trypan Blue Staining Analysis / Counting in Progress Interface



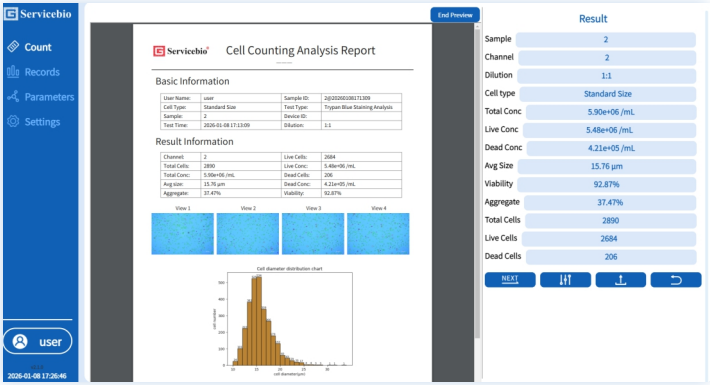
Trypan Blue Staining Analysis/ Counting Results Display Interface



Trypan Blue Staining Analysis/ Calculator Interface



Trypan Blue Staining Analysis/ Cell Diameter Distribution Chart Interface



Trypan Blue Staining Analysis/ Analysis Report Interface

After analysis is complete, the Sample Data column on the right side of the main interface displays the current test results. Click the buttons 1 2 3 4 below the field of view image to switch and view the field of view images.

Click the buttons Channel 1 Channel 2 Channel 3 Channel 4 Channel 5 Channel 6 at the bottom of the main image window to switch and view images from 6 different channels;

Click the button to restore the image to its original centered position with one click;

Click the button to switch between the original image and the result image;

Click the button to view the cell diameter distribution chart;

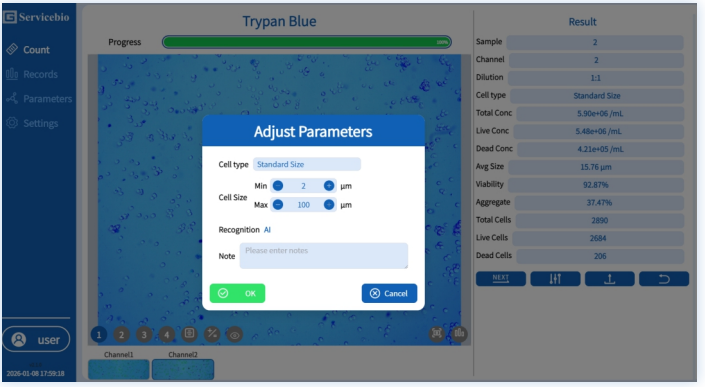
Click the button to pop up the cell passage calculator window, calculating the volume of cell sample and diluent needed to dilute the current sample to a specific concentration;

Click the button Total to select the current sample data type of total cell concentration, live cell concentration, and dead cell concentration for calculation;

Click the button E+03mL to select different concentration orders of magnitude;

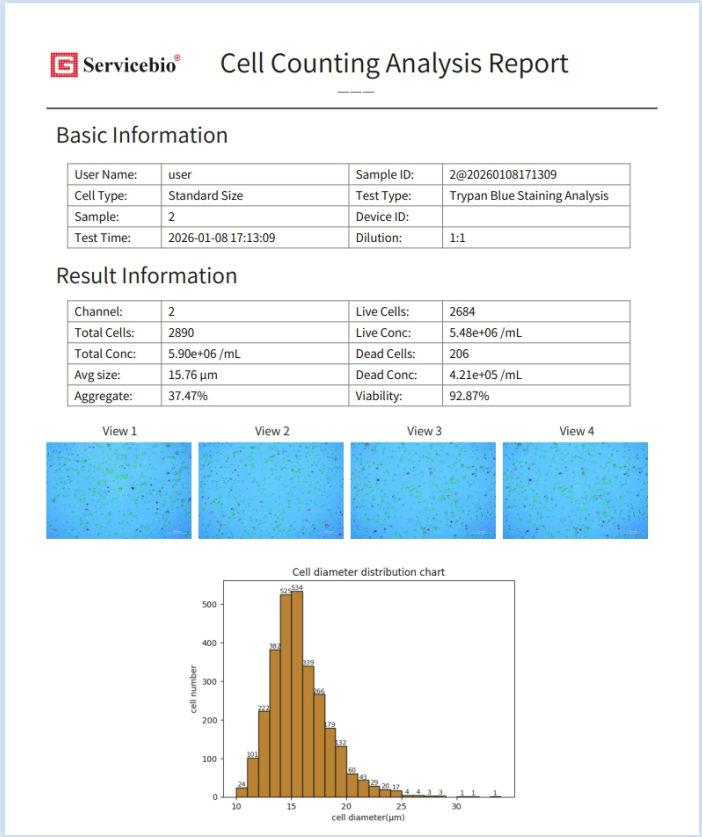
Click the button to export the analysis report when the user needs to output the current sample data;

Click the button to preview the current counting analysis report. The analysis report mainly includes basic information, result information, cell diameter distribution chart, and other data.



Trypan Blue Staining Analysis / Adjust Parameters Interface

After sample counting is complete, click the button to adjust the cell diameter parameters for the current sample data, filter out cells within the specified parameter range, generate an adjustment record, and save it automatically.



Servicebio® Cell Counting Analysis Report

Basic Information

User Name:	user	Sample ID:	2@20260108171309
Cell Type:	Standard Size	Test Type:	Trypan Blue Staining Analysis
Sample:	2	Device ID:	
Test Time:	2026-01-08 17:13:09	Dilution:	1:1

Result Information

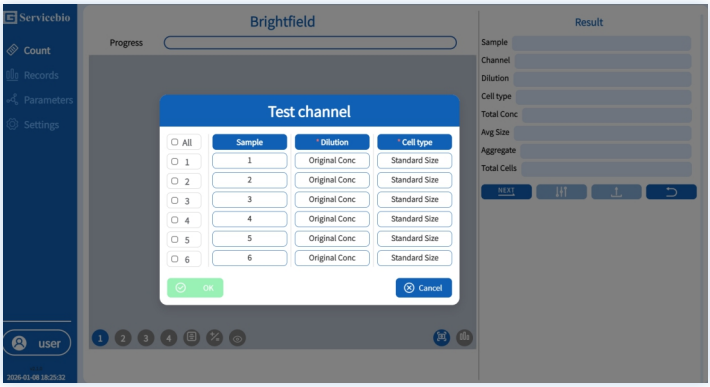
Channel:	2	Live Cells:	2684
Total Cells:	2890	Live Conc:	5.48e+06 /mL
Total Conc:	5.90e+06 /mL	Dead Cells:	206
Avg size:	15.76 μm	Dead Conc:	4.21e+05 /mL
Aggregate:	37.47%	Viability:	92.87%

View 1 View 2 View 3 View 4

Trypan Blue Staining Analysis Report

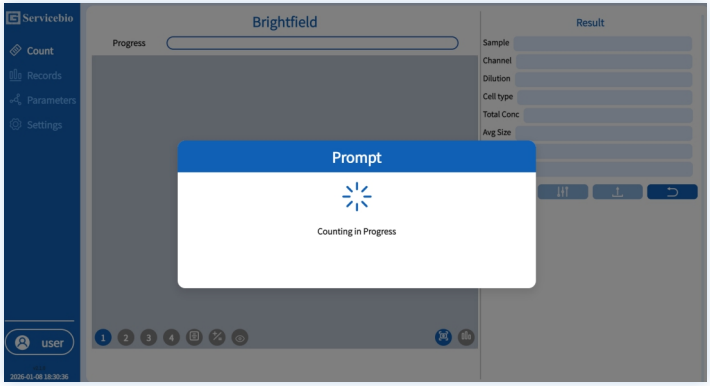
4.4.2 Unstained Analysis

Select the sample measurement function, click the Unstained Analysis mode on the main interface. The main window will pop up the test well selection interface for Unstained Analysis mode. Click "Select All" to simultaneously select all 6 channels for testing, or select a single channel individually for counting analysis. Choose or create a sample name as needed (default sample name is the channel number). Select the dilution ratio (default is 1:1). Select the cell type (default cell type is standard particles).

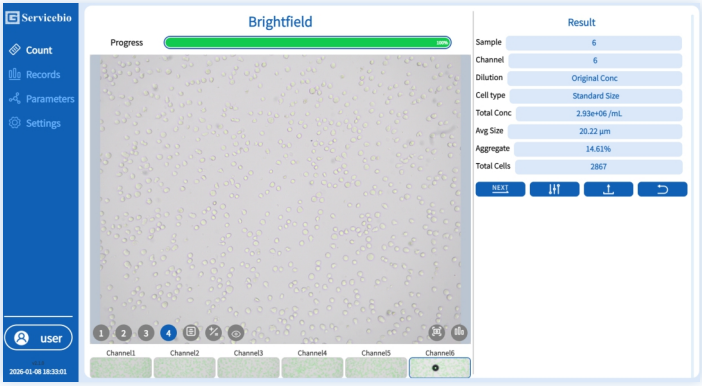


Select Test Well Positions Interface

After selecting the well positions and editing the relevant parameters, click the OK button to start the Unstained Analysis. A "Counting in progress" prompt box will pop up on the interface. Do not perform any operations at this time; wait quietly for the counting to complete.

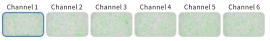


Unstained Analysis / Counting in Progress Interface



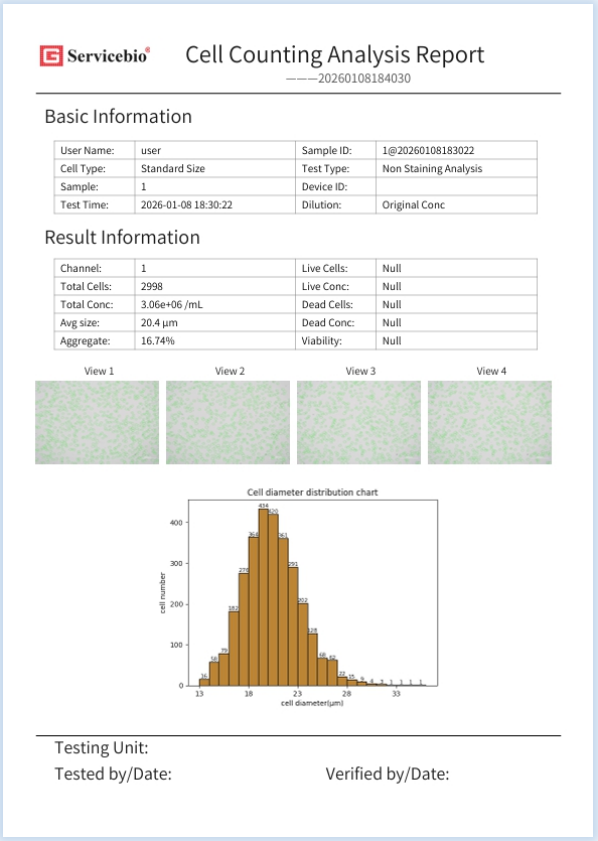
Unstained Analysis / Counting Results Display Interface

After analysis is complete, the Sample Data column on the right side of the main interface displays the current test results. Click the buttons 1 2 3 4 below the field of view image to switch and view the field of view images;

Click the buttons 

at the bottom of the main image window to switch and view images from 6 different channels.

Other interface operation methods are the same as for Trypan Blue Staining Analysis. Refer to section 4.4.1 for details.



Servicebio Cell Counting Analysis Report

———20260108184030

Basic Information

User Name:	user	Sample ID:	1@20260108183022
Cell Type:	Standard Size	Test Type:	Non Staining Analysis
Sample:	1	Device ID:	
Test Time:	2026-01-08 18:30:22	Dilution:	Original Conc

Result Information

Channel:	1	Live Cells:	Null
Total Cells:	2998	Live Conc:	Null
Total Conc:	3.06e+06 /mL	Dead Cells:	Null
Avg size:	20.4 μm	Dead Conc:	Null
Aggregate:	16.74%	Viability:	Null

View 1 View 2 View 3 View 4

Testing Unit:

Tested by/Date: Verified by/Date:

Unstained Analysis Report

4.4.3 AO/PI Staining Analysis

Select the sample measurement function, click the AO/PI Staining Analysis mode on the main interface. The main window will pop up the test well selection interface for AO/PI Staining Analysis mode. Click "Select All" to simultaneously select all 6 channels for testing, or select a single channel individually for counting analysis. Choose or create a sample name as needed (default sample name is the channel number). Select the dilution ratio (default is 1:1). Select the cell type (default cell type is standard particles). Editing methods are detailed in section 4.4.1 Trypan Blue Staining Analysis.

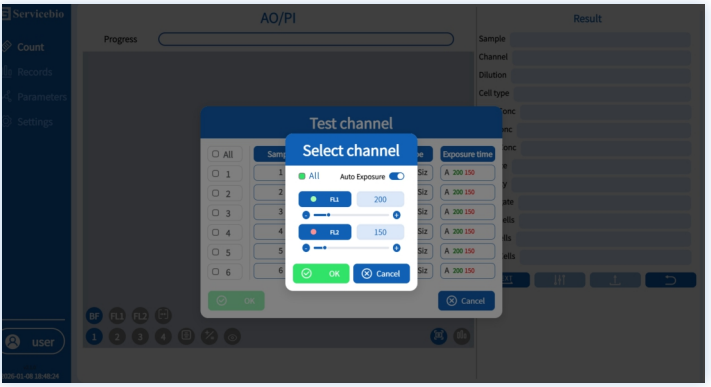
Dual Fluorescence Channel Exposure Settings

Click the "AO/PI Staining Analysis" mode in the sample measurement interface to enter the "Select Test Well Positions" window. Set the exposure parameters in the channel settings. Click the button A 200 150 below the channel, turn off the "Auto Exposure" button, then you can adjust the exposure time for "FL1 (Green Fluorescence)" and "FL2 (Red Fluorescence)". When adjusting parameters, click the corresponding fluorescence channel button ("FL1" or "FL2"), drag the exposure time slider, or manually input the exposure time. If you do not modify the exposure time, click the "Auto Exposure" button. The displayed parameters at this time are the default exposure times: "FL1" default is 200 ms, "FL2" default is 150 ms. These parameters are matched with our dedicated AO/PI staining reagent. Users can fine-tune the exposure parameters based on the actual sample detection situation.

AO/PI Reagent Staining Method

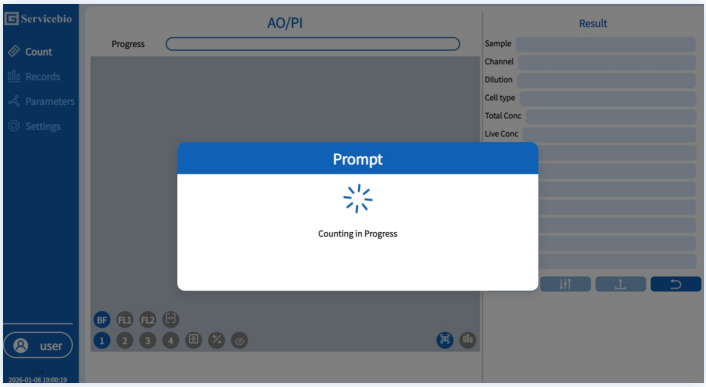
The AO/PI Cell Viability Assay Kit (Fluorescence Method) is a premixed optimal ratio of Acridine Orange (AO) staining solution and Propidium Iodide (PI) staining solution. It is a kit for rapid and convenient detection of cell viability based on double-staining of nuclei with DNA probes. AO is membrane-permeable and can cross the cell membrane to stain DNA and RNA, emitting green fluorescence. PI is a non-cell-membrane-permeable fluorescent dye that cannot cross the intact plasma membrane of viable cells; it can only enter dead or membrane-damaged cells and intercalate into the DNA double helix of dead cells to form a PI-DNA complex, producing red fluorescence, thus distinguishing live and dead cells.

This kit provides a Ready-to-Use 2× AO/PI Fluorescence Dual Stain Reagent. Mix the sample to be tested with the AO/PI dual stain reagent at a 1:1 ratio. It's simple and convenient, no need to prepare and dilute various solutions, eliminating operational errors during preparation and dilution, improving detection accuracy. It can be detected using instruments such as fluorescence microscopes, flow analyzers, and fluorescence cell counters.

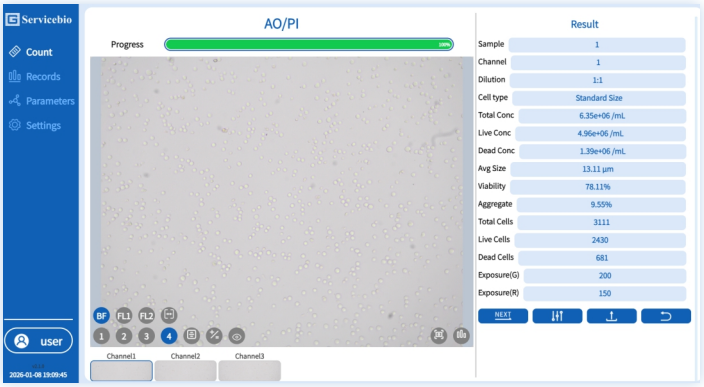


AO/PI Staining Analysis / Select Channel Interface

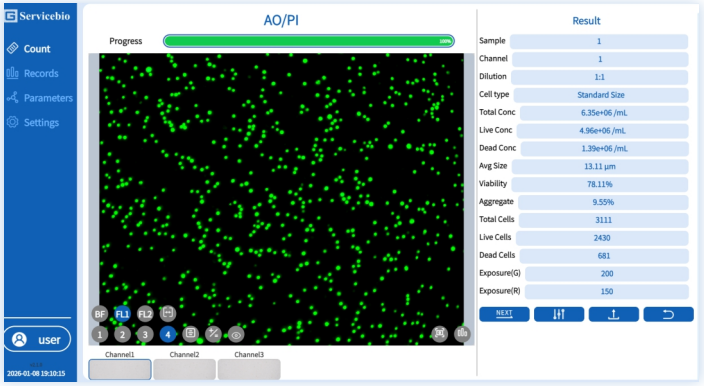
Cell samples stained with the matching reagent can be directly loaded for detection. After parameter settings are complete, click "OK" to start AO/PI staining counting. A "Counting in progress" prompt box will pop up on the screen. Do not perform any operations during the counting process; wait for the counting to finish before viewing the test results.



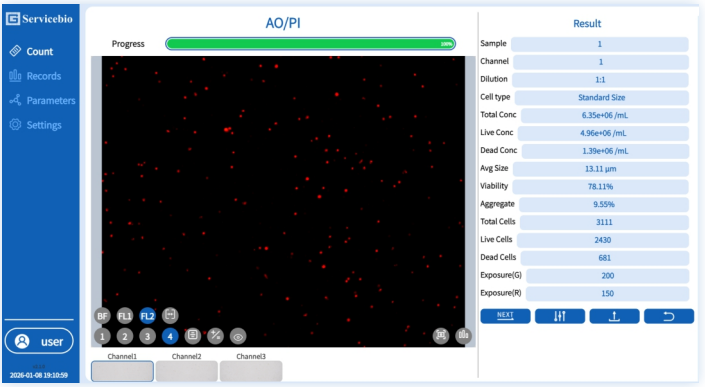
AO/PI Staining Analysis / Counting in Progress Interface



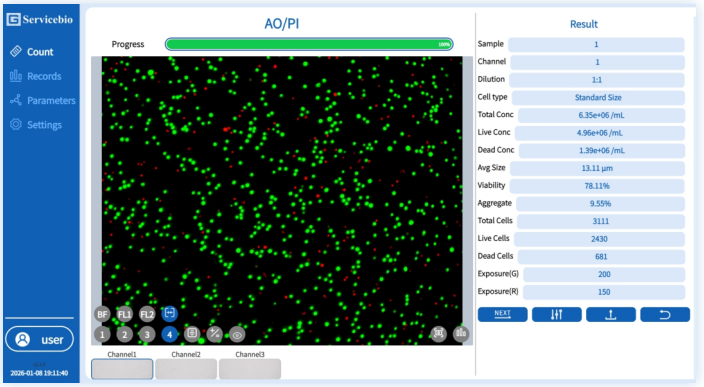
AO/PI Staining Analysis Interface / Counting Results Display Interface (Brightfield Original Image)



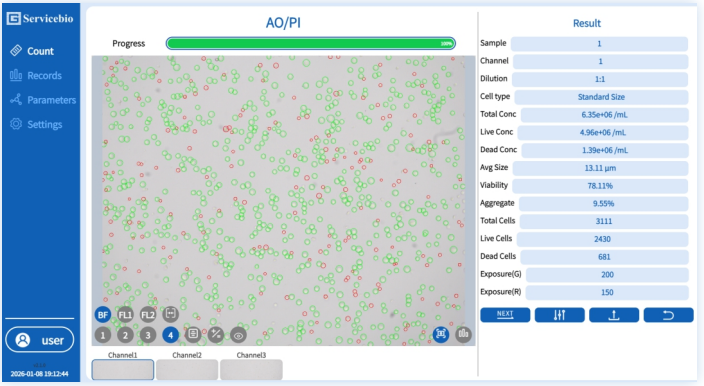
AO/PI Staining Analysis Interface / Counting Results Display Interface (Green Fluorescence Original Image)



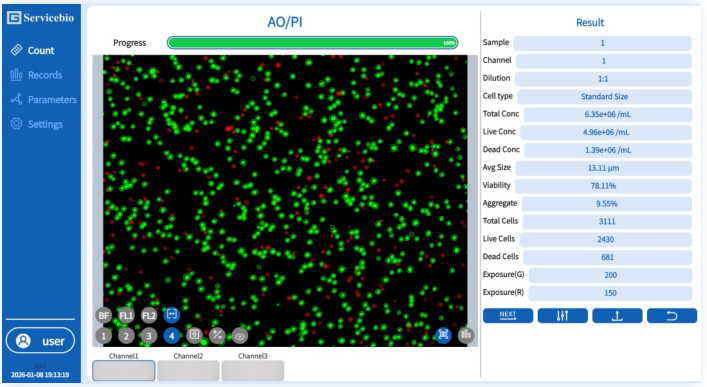
AO/PI Staining Analysis Interface / Counting Results Display Interface (Red Fluorescence Original Image)



AO/PI Staining Analysis Interface / Counting Results Display Interface (Red-Green Fusion Original Image)

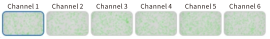


AO/PI Staining Analysis Interface / Counting Results Display Interface (Brightfield Result Image)



AO/PI Staining Analysis Interface / Counting Results Display Interface (Red-Green Fusion Result Image)

After analysis is complete, the Sample Data column on the right side of the main interface displays the current test results. Click the buttons 1 2 3 4 below the field of view image to switch and view the field of view images.

Click the buttons 

at the bottom of the main image window to switch and view images from 6 different channels;

Click the button BL to view the brightfield image;

Click the button FL1 to view the green fluorescence image;

Click the button FL2 to view the red fluorescence image;

Click the button to view the red-green fusion image;

Click the button to restore the image to its original centered position with one click;

Click the button to switch between the original image and the result image;

Click the button to view the cell diameter distribution chart, including brightfield, FL1, and FL2 diameter distribution;

Click the button to pop up the cell passage calculator window, calculating the volume of cell sample and diluent needed to dilute the current sample to a specific concentration;

Click the button Total to select the current sample data type of total cell concentration, live cell concentration, and dead cell concentration for calculation;

Click the button E+03mL to select different concentration orders of magnitude;

Click the button to export the analysis report when the user needs to output the current sample data;

Click the button to preview the current counting analysis report. The analysis report mainly includes basic information, result information, cell diameter distribution chart, and other data;

After sample counting is complete, click the button to adjust parameters such as cell diameter and FL1, FL2 exposure time for the current sample data, filter out cells within the specified parameter range, generate an adjustment record, and save it automatically;

Click the button NEXT proceed to the next counting.

AO/PI Staining Analysis / Adjust Parameters Interface

Servicebio® Cell Counting Analysis Report

———20260108191709

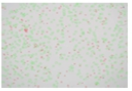
Basic Information

User Name:	user	Sample ID:	1@20260108190503
Cell Type:	Standard Size	Test Type:	AO/PI Staining Analysis
Sample:	1	Device ID:	
Test Time:	2026-01-08 19:05:03	Dilution:	1:1

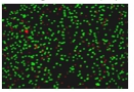
Result Information

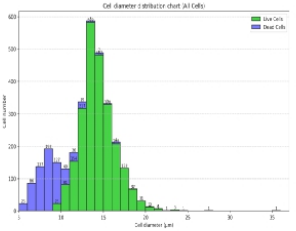
Channel:	1	Live Cells:	2430
Total Cells:	3111	Live Conc:	4.96e+06 /mL
Total Conc:	6.35e+06 /mL	Dead Cells:	681
Avg size:	13.11 μm	Dead Conc:	1.39e+06 /mL
Aggregate:	9.55%	Viability:	78.11%
Exposure(G):	200ms	Exposure(R):	150ms

BF Result Display



FL Merge Result Display





Testing Unit:

Tested by/Date: Verified by/Date:

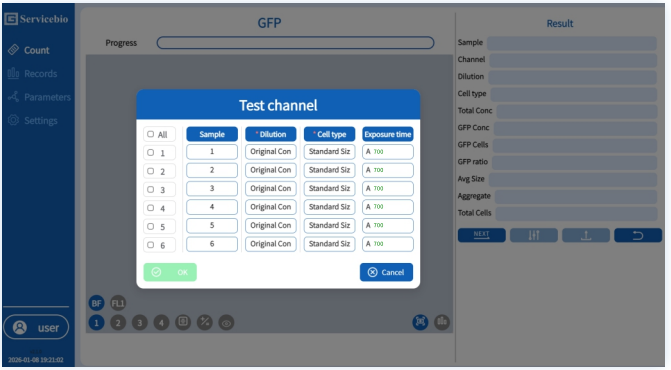
AO/PI Staining Analysis Report

4.4.4 GFP Transfection Analysis

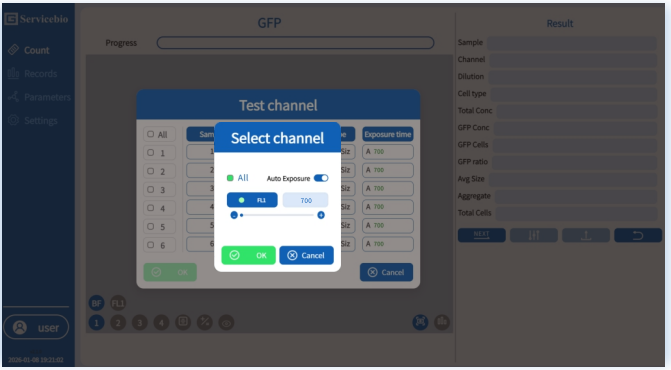
Select the sample measurement function, click the GFP Transfection Analysis mode on the main interface. The main window will pop up the test well selection interface for GFP Transfection Analysis mode. Click "Select All" to simultaneously select all 6 channels for testing, or select a single channel individually for counting analysis. Choose or create a sample name as needed (default sample name is the channel number). Select the dilution ratio (default is 1:1). Select the cell type (default cell type is standard particles). Editing methods are detailed in section 4.4.1 Trypan Blue Staining Analysis.

GFP Exposure Settings

Click the "GFP Staining Analysis" mode in the sample measurement interface to enter the "Select Test Wells" window. Set the exposure parameters in the channel settings. Click the A 700 button below the channel, turn off the "Auto Exposure" button, then you can adjust the exposure time for "FL1 (Green Fluorescence)". When adjusting parameters, click the "FL1" fluorescence channel button, drag the exposure time slider, or manually input the exposure time. If you do not modify the exposure time, click the "Auto Exposure" button. The displayed parameters at this time are the default parameters. You can adjust the current counting status. The default exposure time for "FL1" is 700 ms, which is the default parameter. Users can fine-tune the exposure parameters based on the actual sample detection situation.

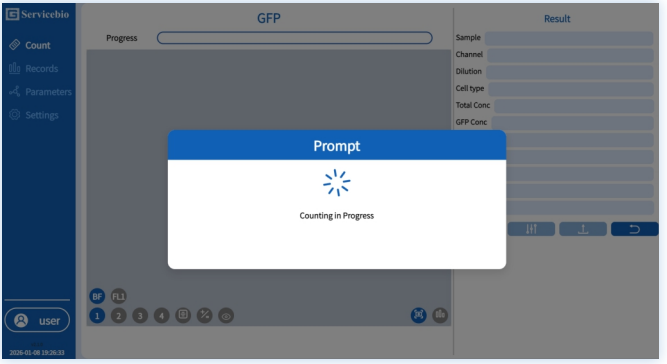


GFP Transfection Analysis / Select Test Well Positions Interface

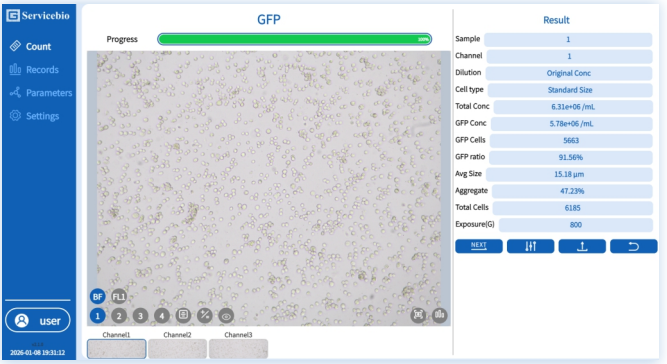


GFP Transfection Analysis / Select Channel Interface

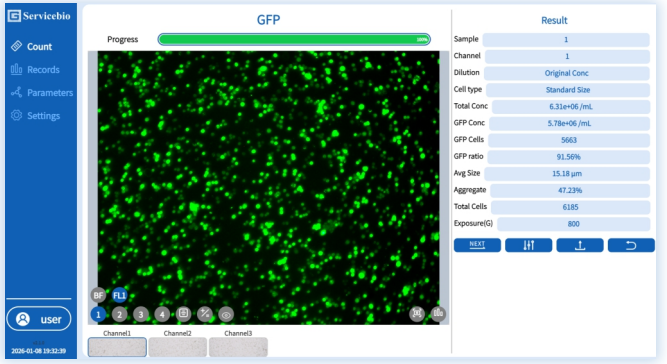
Cell samples stained with the matching reagent can be directly loaded for detection. After parameter settings are complete, click "OK" to start GFP Transfection Analysis. A "Counting in progress" prompt box will pop up on the screen. Do not perform any operations during the counting process; wait for the counting to finish before viewing the test results.



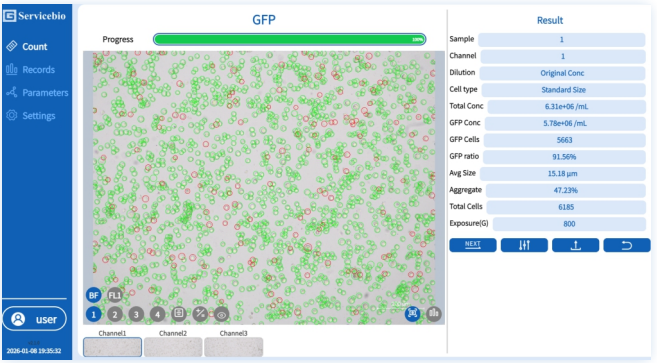
GFP Transfection Analysis / Counting in Progress Interface



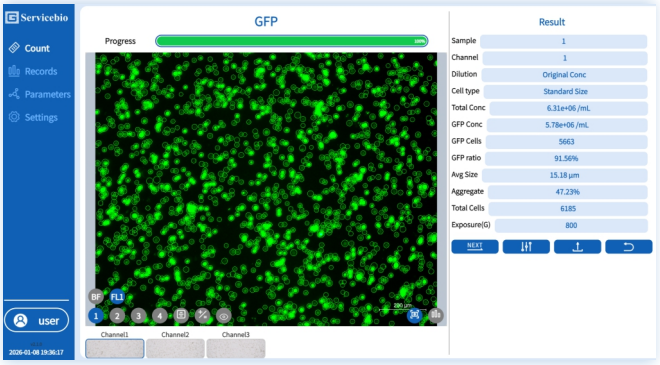
GFP Transfection Analysis Interface (Brightfield Original Image)



GFP Transfection Analysis Interface (Green Fluorescence Original Image)

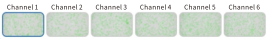


GFP Transfection Analysis Interface (Brightfield Result Image)



GFP Transfection Analysis Interface (Green Fluorescence Result Image)

After analysis is complete, the Sample Data column on the right side of the main interface displays the current test results. Click the buttons 1 2 3 4 below the field of view image to switch and view the field of view images.

Click the buttons 

at the bottom of the main image window to switch and view images from 6 different channels;

Click the button BL to view the brightfield image;

Click the button FL1 to view the green fluorescence image;

Click the button to restore the image to its original centered position with one click;

Click the button to switch between the original image and the result image;

Click the button to view the cell diameter distribution chart, including brightfield and FL1 diameter distribution;

Click the button to pop up the cell passage calculator window, calculating the volume of cell sample and diluent needed to dilute the current sample to a specific concentration;

Click the button Total to select the current sample data type of total cell concentration, live cell concentration, and dead cell concentration for calculation;

Click the button E+03mL to select different concentration orders of magnitude;

Click the button to export the analysis report when the user needs to output the current sample data.;

Click the button to preview the current counting analysis report. The analysis report mainly includes basic information, result information, cell diameter distribution chart, and other data;

After sample counting is complete, click the button to adjust parameters such as cell diameter and FL1, FL2 exposure time for the current sample data, filter out cells within the specified parameter range, generate an adjustment record, and save it automatically.;

Click the button NEXT to proceed to the next counting;

Click the button to restore the image to its original centered position with one click.;

Click the button to switch between the original image and the result image.;

Click the button to view the cell diameter distribution chart.;

Click the button to pop up the cell passage calculator window, calculating the volume of cell sample and diluent needed to dilute the current sample to a specific concentration.;

Click the button Total to select the current sample data type of total cell concentration, live cell concentration, and dead cell concentration for calculation.;

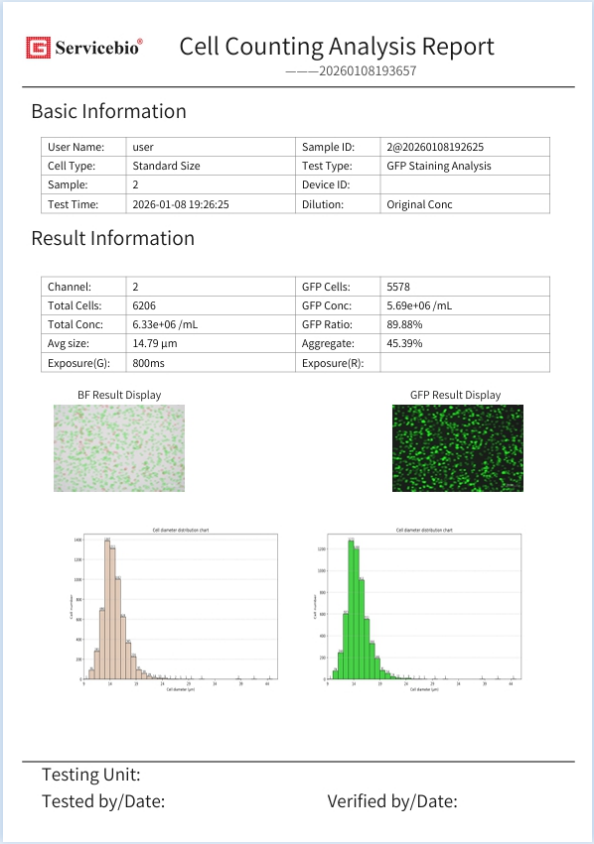
Click the button E+03mL to select different concentration orders of magnitude.;

Click the button to export the analysis report when the user needs to output the current sample data.;

Click the button to preview the current counting analysis report. The analysis report mainly includes basic information, result information, cell diameter distribution chart, and other data.;

After sample counting is complete, click the button to adjust parameters of cell diameter, filter out cells within the specified parameter range, generate an adjustment record, and save it automatically.;

Click the button NEXT to proceed to the next counting.


Servicebio

Cell Counting Analysis Report

——20260108193657

Basic Information

User Name:	user	Sample ID:	2@20260108192625
Cell Type:	Standard Size	Test Type:	GFP Staining Analysis
Sample:	2	Device ID:	
Test Time:	2026-01-08 19:26:25	Dilution:	Original Conc

Result Information

Channel:	2	GFP Cells:	5578
Total Cells:	6206	GFP Conc:	5.69e+06 /mL
Total Conc:	6.33e+06 /mL	GFP Ratio:	89.88%
Avg size:	14.79 μm	Aggregate:	45.39%
Exposure(G):	800ms	Exposure(R):	

BF Result Display

GFP Result Display

Testing Unit:

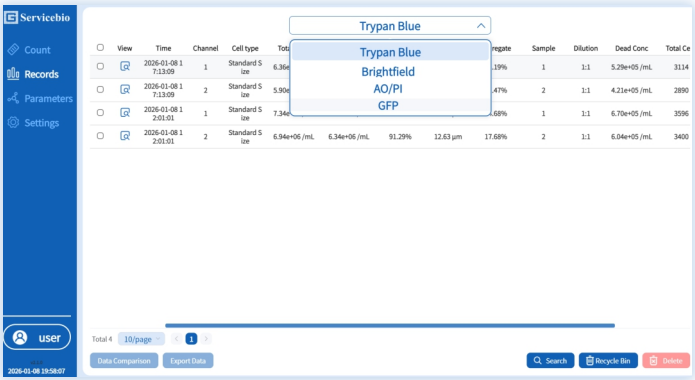
Tested by/Date:

Verified by/Date:

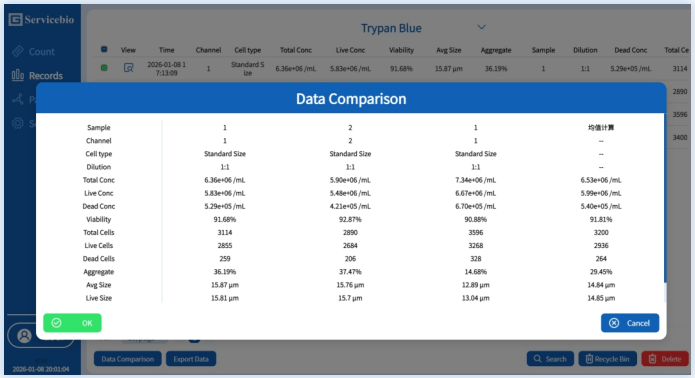
GFP Transfection Analysis Report

4.5 Test Records

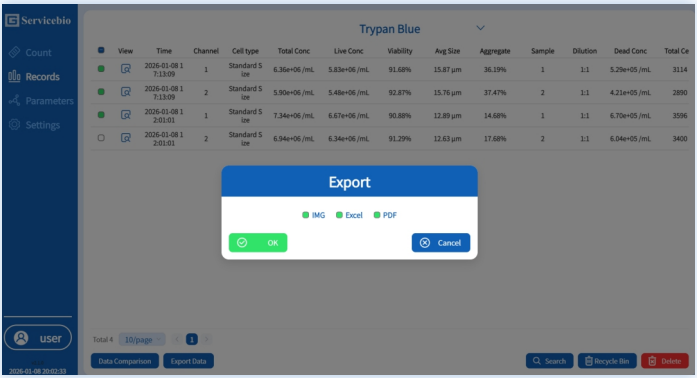
Click the button Records on the left side of the main interface to enter the test records interface. Click the "Trypan Blue Staining Analysis" text at the top of the main interface to pop up a selection box for four different analysis modes: Trypan Blue Staining Analysis / Unstained Analysis / AO/PI Staining Analysis / GFP Transfection Analysis. Click the corresponding analysis mode to enter its test records interface.



Test Records Interface



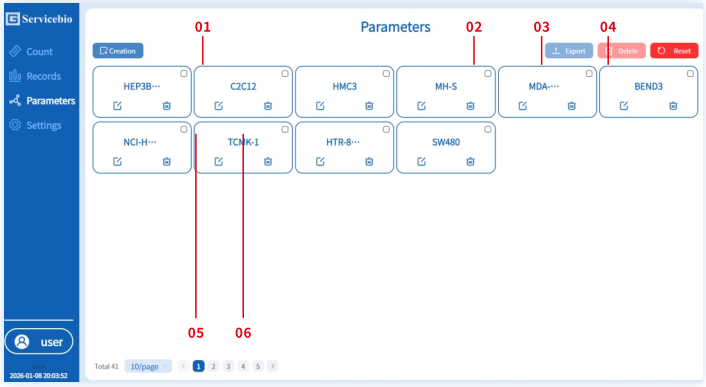
Data Comparasion Interface



Data Export Interface

Click the button Records on the left side of the main interface to view, compare data, and export test records for each analysis module: Trypan Blue Staining Analysis, Unstained Analysis, AO/PI Staining Analysis, and GFP Transfection Analysis.

4.6 Cell Parameters



Cell Parameters Interface

01 **New** Create new cell parameters

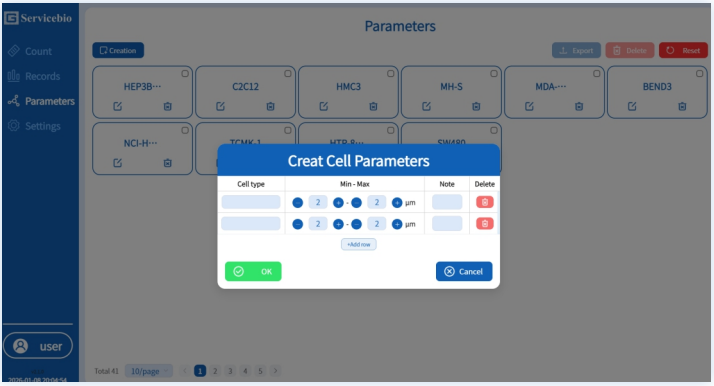
02 **Export** Export cell parameter records

03 **Delete** Batch delete cell parameters

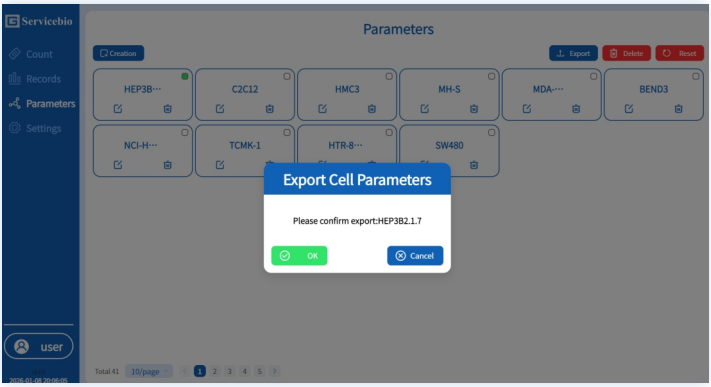
04 **Reset** Reset data in cell parameters

05 **Edit** Edit these cell parameters

06 **Delete** Delete these cell parameters



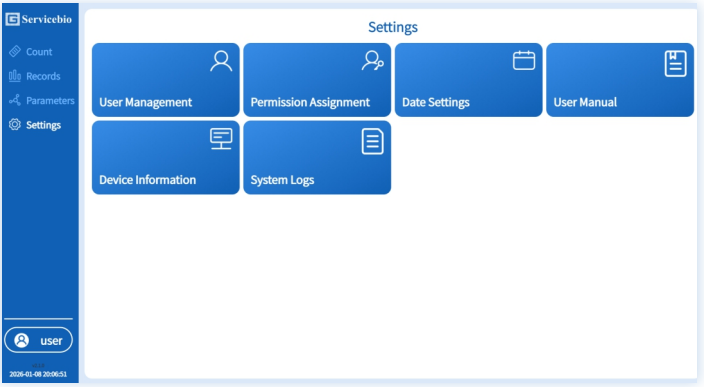
Create New Cell Parameters Interface



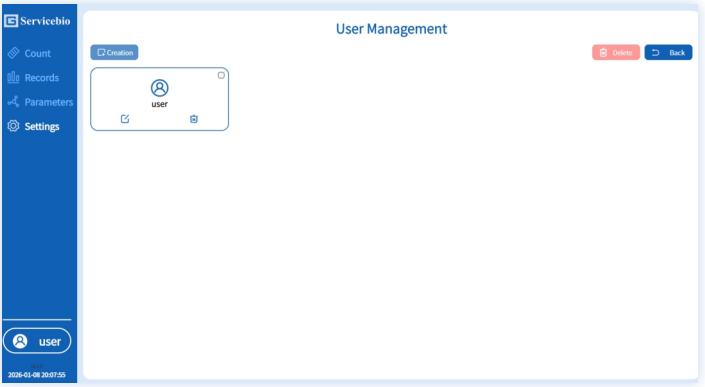
Cell Parameters Export Interface

Click the Cell Parameters window in the left navigation bar to enter the cell parameters interface. Click Creation, fill in the cell type and cell diameter range to create new cell parameters.

4.7 System Settings



System Settings Interface



User Management Interface

01 **New** Create new accout

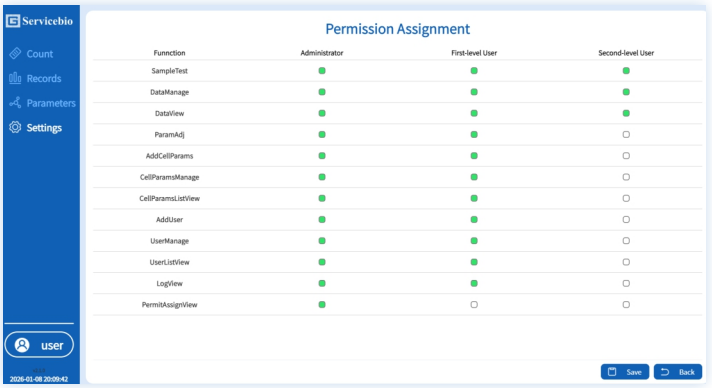
02 **Delete** Batch delete accounts

03 **Back** Back to previous interface

04 **Edit** Edit this account

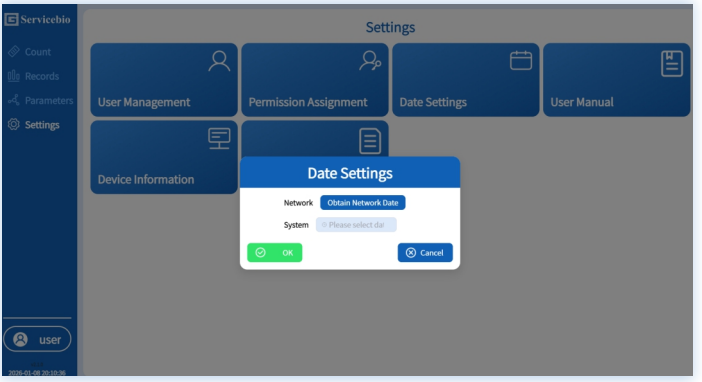
05 **Delete** Delete this account

Click  to enter the user management interface.



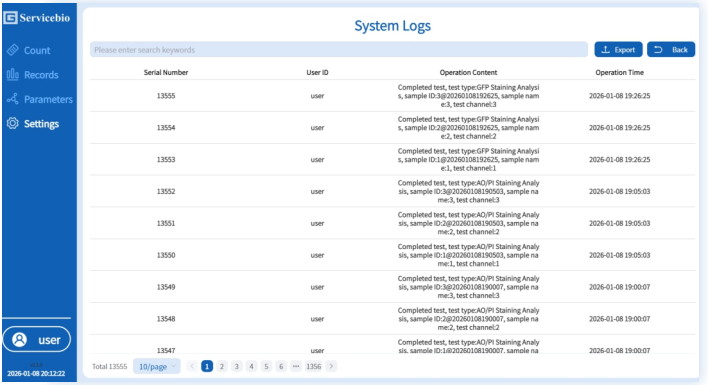
Permission Assignment

Click  to enter the permission assignment interface.



Date Settings Interface

Click  to enter the date settings interface.



System Logs Interface

Click  to enter the system logs interface.

4.8 Cell Counting

1. Take 10μL of the prepared sample and add it to the loading area of the cell counting chamber;

2. After sample loading is complete, horizontally insert the cell counting chamber into the port at the front of the fluorescence cell counter;

3. Perform the pre-counting setup operations on the fluorescence cell counter;

4. Start cell counting. After counting is finished, the sample data column on the right side of the screen displays the cell counting results;

5. The main window in the center of the screen displays cell images (original and result images), channel numbers, and field numbers. There are 6 channels in total, each with four images.

4.9 Shutdown Procedure

The user can click the button in the lower left corner to perform the shutdown operation.

05 Warranty Service

Warranty Service Description

If the instrument or parts are damaged during the warranty period, we will be responsible for free repair or replacement of the damaged components.

Damage caused by the following reasons is excluded:

1. Damage caused by improper use;

2. Repair or modification not carried out by our company;

3. Replacement of parts not produced or authorized by our company;

4. Contamination and corrosion caused by improper reagents, solvents, or samples.

If you require more services, please visit our official website or call our national service hotline: 400-6027178

When purchasing the product, please fill in the following warranty card information carefully and keep it properly.

Product Name	
Cat.No.	
Purchase Date	
Address	
Model	
Product Number	
Quality Feedback	

Servicebio

Wuhan Servicebio Technology Co., Ltd.

400-6027-178

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